

## LA Hot Start Plain Master Mix

(Catalogue number L611, L612, L613, L613xl)

rev. 04/2025

### Description

LA Hot Start Plain Master Mix is optimized mix for universal use in PCR. It contains a blend of two DNA polymerases, deoxyribonucleotide triphosphates (dNTPs), anti-Taq monoclonal antibody, buffer, stabilizers, additives and other components. **This product is the most advanced variant of Master mix for PCR:**

### **Rapid samples preparation, 500 µl aliquots**

- 2x concentrated Master Mix is in 500 µl aliquots. This decreases possibility of contamination and allows rapid preparation of the samples by adding just specific oligonucleotide primers, template DNA and H<sub>2</sub>O (included).

### **LA (Long and Accurate)**

- The Mix contains unique blend of two thermostable DNA polymerases. One of them is Taq DNA polymerase, which is highly processive but lacks a 3'→5' exonuclease proofreading activity, which accounts for relatively high error rate during DNA amplification. Second polymerase is less processive but possesses 3'→5' exonuclease activity necessary for proofreading. The polymerases mix decreases error rate during synthesis of complementary DNA strands and preserves high speed of DNA synthesis. The repair capability allows amplification of complex genomic DNA fragments (up to 15 kbps in size) and less complex viral DNA (up to 40 kbps in size) and significantly decreases error rate when compared to Taq DNA polymerase

### **Hot Start**

- The Mix contains monoclonal antibody, which binds DNA polymerases and thereby blocks their enzymatic activity and amplification of nonspecific DNA fragments. After the first denaturation cycle, the antibody is irreversibly heat-inactivated and Taq DNA polymerase regains enzymatic activity.

### **Suitable for real-time PCR (qPCR)**

- The Mix does not contain a dye and therefore before electrophoresis it is necessary to add loading buffer.
- Because of absence of a dye the Mix can be also used for qPCR.

### **High efficiency and specificity**

- The Mix allows highly sensitive and specific amplification of corresponding fragments of DNA or cDNA; it possesses MgCl<sub>2</sub> at a concentration suitable for most PCRs.

## Technical data

### **Components and packaging**

- 1 tube with 0.5 ml LA Hot Start Plain Master Mix (for 40 reactions, 25 µl each).
- 1 tube with 1.5 ml Ultra PCR H<sub>2</sub>O.

### **Composition**

- 2x concentrated LA Hot Start Plain Master Mix contains: 150 mM Tris-HCl, pH 8.8 (at 25°C), 40 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.02% Tween 20, 5 mM MgCl<sub>2</sub>, 400 µM dATP, 400 µM dCTP, 400 µM dGTP, 400 µM dTTP, 100 U/ml Taq DNA polymerase blend, monoclonal antibody anti-Taq (38 nM), stabilizers and additives.

### **Storage**

- At temperature -20°C ± 5°C; shortly (up to one week) at temperature 4 - 30°C. This decreases demands for transportation (Nature friendly). Material can be repeatedly defrosted.

### **Purity and quality control**

- Purity of Taq DNA polymerases is verified electrophoretically (SDS PAGE). Material is free of nucleases.
- Each batch of LA Hot Start Plain Master Mix is tested for amplification of a single copy gene in genomic DNA.

## Protocol

### Suggested basic protocol for PCR amplification using LA Hot Start Master Mix-Plain

1. In a thin-walled PCR tube the following components are mixed:

Volume*	Reagent	Final concentration
12.5 µl	LA Hot Start Master Mix-Plain	1x LA Hot Start Master Mix-Plain (75 mM Tris-HCl, pH 8.8, 20 mM (NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub> , 0.01% Tween 20, 2.5 mM MgCl <sub>2</sub> , 200 µM dATP, 200 µM dCTP, 200 µM dGTP, 200 µM dTTP, 50 U/ml blend of DNA polymerases, 19 nM anti-Taq monoclonal antibody)
1 µl	5' primer	0.1 - 1 µM (~ 20 bases)
1 µl	3' primer	0.1 - 1 µM (~ 20 bases)
1 µl	Template DNA	
9.5 µl	PCR H <sub>2</sub> O	to a final volume 25 µl

\*Different volumes can be used, but Master Mix should be finally diluted twice.

2. Mix gently and briefly centrifuge.

3. Add ~20 µl of PCR oil (Cat. No. P043) to prevent evaporation (this is not required if thermal cycler with a heated lid is used).

4. Perform PCR under conditions optimized for the primers used. Common cycling parameters are:

	Temperature	Time	Number of cycles
Initial denaturation	94°C	1 min	1
Denaturation	94°C	15 s	25-35
Primers annealing	55-68°C <sup>1</sup>	15 s	
Extension	72°C	1 min per 1 kb	
Final extension	72°C	7 min	1
Cooling	22°C		

<sup>1</sup> Should be determined experimentally; usually 5°C below melting temperature of the primers.

Cat. No	Product name and specification	Quantity
L611	LA Hot Start Plain Master Mix	40 reactions
L612	LA Hot Start Plain Master Mix	200 reactions
L613	LA Hot Start Plain Master Mix	1000 reactions
L613xl	LA Hot Start Plain Master Mix	4x 1000 reactions

